



**JAI HIND COLLEGE
BASANTSING INSTITUTE OF SCIENCE
&
J.T.LALVANI COLLEGE OF COMMERCE
(AUTONOMOUS)**

"A" Road, Churchgate, Mumbai - 400 020, India.

**Affiliated to
University of Mumbai**

Program : B.Sc

Proposed Course : Microbiology T.Y.B.Sc Sem VI

**Credit Based Semester and Grading System (CBGS) with
effect from the academic year 2019-20**

T.Y.B.Sc. Microbiology Syllabus

Academic year 2019-2020

Semester VI			
Course Code	Course Title	Credits	Lectures /Week
SMIC601	rDNA TECHNOLOGY, BIOINFORMATICS & VIROLOGY	2.5	04
UNIT 1	Recombinant DNA Technology		
UNIT 2	Applications of rDNA Technology & Bioinformatics		
UNIT 3	Regulation & Basic Virology		
Unit 4	Advanced Virology		
SMIC602	MEDICAL MICROBIOLOGY AND IMMUNOLOGY PART-II	2.5	04
UNIT 1	Study of vector borne, sexually transmitted and CNS infections		
UNIT 2	Chemotherapy of infectious agents		
UNIT 3	Immunology -I		
Unit 4	Immunology -II		
SMIC603	MICROBIAL BIOCHEMISTRY: PART-II	2.5	04
UNIT 1	Lipid Metabolism & Catabolism of Hydrocarbons		
UNIT 2	Metabolism of Proteins and Nucleic Acids		
UNIT 3	Metabolic Regulation		
Unit 4	Prokaryotic Photosynthesis & Inorganic Metabolism		
SMIC604	BIOPROCESS TECHNOLOGY- PART-II	2.5	04
UNIT 1	Advances in bioprocess technology		
UNIT 2	Pharmaceutical microbiology		
UNIT 3	Instrumentation and IPR		
Unit 4	Industrial fermentations		
SMIC6PR1	Practical	03	08
SMIC6PR2	Practical	03	08

Course Code SMIC601	Course Title: rDNA TECHNOLOGY, BIOINFORMATICS & VIROLOGY (Credits:2.5 Lectures/Week: 04)	
	<p>Learning Objectives:</p> <ul style="list-style-type: none"> ➤ To study recombinant DNA technology and its applications ➤ To understand plasmid and transposons and their importance ➤ To know role of bioinformatics in biology ➤ To learn about Viruses <p>Learning Outcomes: On completion of this course students will learn about the recombinant DNA technology, use of bioinformatic tools and viruses.</p>	
Unit I	Recombinant DNA Technology	15 L
1.1	Model Organisms	01
1.1.1	Characteristics of a model organism	
1.1.2	Examples of model organisms used in study	
1.2	Plasmids	02
1.2.1	Physical nature	
1.2.2	Detection and isolation of plasmids	
1.2.3	Plasmid incompatibility and Plasmid curing	
1.2.4	Cell to cell transfer of plasmids	
1.2.5	Types of plasmids	
1.2.6	Resistance Plasmids, Plasmids encoding Toxins and other virulence characteristics, Col factor, Degradative plasmids	
1.3	Transposable Elements in Prokaryotes	
1.3.1	Insertion sequences	
1.3.2	Transposons: Types, Structure and properties, Mechanism of transposition, Integrons	02
1.4	Basic Steps in Gene Cloning	01
1.5	Cutting and joining DNA molecules	
	Restriction and modification systems, restriction endonucleases, DNA ligases	03

1.6	Vectors	
1.6.1	Plasmids as cloning vectors. plasmid vectors, pBR322 vector	03
1.6.2	Cloning genes into pBR322	
1.6.3	Phage as cloning vectors, cloning genes into phage vector	
1.6.4	Cosmids	
1.6.5	Shuttle vectors	
1.6.6	YAC	
1.6.7	BAC	
1.7	Methods of transformation	01
1.8	Screening and selection methods for identification and isolation of recombinant cells	02
Unit II	Applications of rDNA Technology & Bioinformatics	15 L
2.1	PCR -basic PCR and different types of PCR (Reverse transcriptase PCR, Real time quantitative PCR)	02
2.2	Basic techniques	02
2.2.1	Southern, Northern and Western blotting.	
2.2.1	Autoradiography(explain the terms)	
2.3	Applications of recombinant DNA technology: Site specific mutagenesis of DNA, Uses of DNA polymorphism, STRS and VNTRS, DNA molecular testing for human genetic diseases (Only RFLP), DNA typing, gene therapy, Genetic engineering of plants and animals.	06
2.4	Bioinformatics	05
2.4.1	Introduction: Genomics - structural, functional and comparative genomics	
2.4.2	Proteomics- structural and functional proteomics, Annotation, Transcriptomics, Metabolomics, Pharmacogenomics,	
2.4.3	Sequence alignment & all related terms in bioinformatics	
2.4.4	Database, tools and their uses NCBI, Ex PASY proteomics server, EBI	
2.4.5	Importance, Types and classification of databases Nucleic acid sequence databases- EMBL, DDBJ, GenBank Protein sequence databases-PIR, SWISS-PROT Metabolic Databases - KEGG, METACYC BLAST with one example	
Unit III	Regulation & Basic Virology	15 L
3.1	A) Lac operon and problems on Lac operon B)Trp operon	07

3.2	Viral architecture- Capsid, viral genome and envelope	02
3.3	Viral classification (Baltimore classification)	01
3.4	Viral replication cycle- Attachment, penetration, uncoating, types of viral genome, their replication, assembly, maturation & release.	02
3.5	Regulation of lytic and lysogenic pathway of lambda phage	03
Unit IV	Advanced Virology	15 L
4.1	Structure and Lifecycle of TMV, T4, Influenza virus, HIV	05
4.2	Cultivation of viruses- cell culture techniques, embryonated egg, laboratory animals, Inclusion bodies, Cytopathic effects	03
4.3	Visualization and enumeration of virus particles	03
4.3.1	Measurement of infectious units	
4.3.1.1	Plaque assay	
4.3.1.2	Fluorescent focus assay	
4.3.1.3	Infectious center assay	
4.3.1.4	Transformation assay	
4.3.1.5	Endpoint dilution assay.	
4.3.2	Measurement of virus particles and their components	
4.3.2.1	Electron microscopy	
4.3.2.2	Atomic force microscopy	
4.3.2.3	Haemagglutination	
4.3.2.4	Measurement of viral enzyme activity	
4.4	Role of viruses in Cancer: Important definitions, characteristics of cancer cell, Human DNA tumor viruses- EBV, Kaposi's sarcoma virus, Hepatitis B and C virus, Papiloma Virus.	02
4.5	Prions: Definition, Examples of diseases caused by prions, Kuru, PrP protein and protein only hypothesis	01
4.6	Viroids	01
	<p>Text books:</p> <ol style="list-style-type: none"> 1. Peter J. Russell (2006), "Genetics-A molecular approach", 2nd edition. Pearson International 2. Benjamin A. Pierce (2008), "Genetics a conceptual approach", 3rd edition, W. H. Freeman and company. 3. R. H. Tamarin, (2004), "Principles of genetics", Tata McGraw Hill. 4. M. Madigan, J. Martinko, J. Parkar, (2009), "Brock Biology of microorganisms", 12th edition, Pearson Education International. 5. Fairbanks and Anderson, (1999), "Genetics", Wadsworth Publishing Company. 6. Prescott, Harley and Klein, "Microbiology", 7th edition McGraw Hill International edition. 	

7. Edward Wagner and Martinez Hewlett, (2005) "Basic Virology", 2nd edition, Blackwell Publishing
 8. Teri Shors. (2009), "Understanding viruses", Jones and Bartlett publishers.
 9. S. Ignacimuthu, (2005), "Basic Bioinformatics", Narosa publishing house.
 10. Robert Weaver, (2008), "Molecular biology", 3rd edition, McGraw Hill International edition.
 11. Primrose and Twyman, (2001), "Principles of gene manipulation and genomics", 6th edition, Blackwell Publishing
 12. Arthur Lesk, (2009), "Introduction to Bioinformatics", 3rd edition, Oxford University Press
 13. Snustad, Simmons, "Principles of genetics", 3rd edition. John Wiley & sons, Inc.
 14. R.C. Dubey S. Chand. (2010) A textbook of biotechnology 4th edition.
 15. Pelczar, M., Reid, R. and Chan, E. (1986). Microbiology 5th ed. New York: McGraw-Hill
 16. Willey, J. M., Sherwood, L., Wool verton, C. J., Prescott, L. M., & Willey, J. M. (2011). Prescott's microbiology 8th ed. New York: McGraw-Hill
 17. Kindt, Goldsby, Osborne Kuby Immunology, 4th and 6th edition, WH Freeman and Company
- Reference books:**
1. Flint, Enquist, Racanillo and Skalka, "Principles of virology", 2nd edition. ASM press.
 2. T.K. Attwood & D.J. Parry-Smith, (2003), "Introduction to bioinformatics", Pearson education
 3. Benjamin Lewin, "Genes IX", (9th edition), Jones and Bartlett publishers.
 4. JD Watson, "Molecular biology of the gene", 5th edition.

Course Code SMIC602	Course Title: MEDICAL MICROBIOLOGY AND IMMUNOLOGY PART-II	2.5 Credits Lectures/Week 04
	Learning Objectives : <ul style="list-style-type: none"> ➤ To study Vector borne, Sexually transmitted and CNS infections ➤ To understand the principles of Chemotherapy ➤ To learn the role of T and B cells in generating adaptive immunity and study effector responses in both Humoral & Cell Mediated Immunity ➤ To apply the concept of immunity to prevention of disease by development of vaccines ➤ To understand the concepts of immunohaematology, Hypersensitivity and autoimmunity 	
	Learning outcomes : On completion of the course the students will be able to: <ul style="list-style-type: none"> ➤ Comment on the different pathogens causing vector borne, sexually transmitted and CNS infections and the disease caused by them wrt transmission, pathogenesis and clinical manifestation, Lab diagnostic procedures and prophylactic measures. ➤ Describe the mode of action of antibiotics ➤ Understand the role of T and B cells in immunity ➤ Understand the principles and use of Vaccines ➤ Explain the basic principles of immunohaematology, Hypersensitivity and Autoimmunity. 	
	THEORY	(45 lectures)
Sub Unit	Unit – I: Study of vector borne, sexually transmitted and CNS infections: (Few Diseases with Emphasis on Characteristics of the Etiological Agent, Pathogenesis, Laboratory Diagnosis and Prevention only)	15 lectures
1.1	Study of vector-borne infections- Malaria, Leptospirosis	03
1.2 1.2.1 1.2.2 1.2.3	Study of sexually transmitted infectious diseases Syphilis AIDS Infection due to Hepatitis B	07
1.3 1.3.1	Study of central nervous system infectious diseases Tetanus	

1.3.2	Polio	05
1.3.3	Meningococcal meningitis	
1.3.4	Rabies	
Sub Unit	Unit II: Chemotherapy of Infectious Agents	15 lectures
2.1	Attributes of an ideal chemotherapeutic agent- Selective toxicity, Bioavailability of drug routes of drug administration, LD50, MBC, etc	02
2.2	Mode of action of antibiotics on-	07
2.2.1	Cell wall (Beta- lactams- Penicillin and Cephalosporins, Carbapenems, Vancomycin)	
2.2.2	Cell Membrane (Polymyxin and Imidazole)	
2.2.3	Protein Synthesis (Streptomycin, Tetracycline Chloramphenicol and Erythromycin)	
2.2.4	Nucleic acid (Quinolones, Nalidixic acid, Rifamycin)	
2.2.5	Enzyme inhibitors (Sulfa drugs, Trimethoprim)	
2.3	List of common antibiotics –used for treating viral, fungal and parasitic diseases.	01
2.4	Mechanisms of drug resistance- Its evolution, pathways and origin for ESBL, VRE, MRSA	03
2.5	(i) Selection and testing of antibiotics for bacterial isolates by Kirby- Bauer method, E test (ii) Methods that detect <i>S.aureus</i> resistance to methicillin, and determination of ESBL strains	02
Sub Unit	Unit III: Immunology–I	15 Lectures
3.1	T cells	05
3.1.1	T Cell Receptor-structure (alpha-beta, gamma-delta TCR) TCR-CD ₃ complex-structure and functions. Accessory molecules	
3.1.2	Development and maturation of T cells , Thymic Selection of the T –cell Repertoire	
3.1.3	T cell activation	

3.1.3.1	TCR mediated signaling – Overview	
3.1.3.2	Costimulatory signals	
3.1.3.3	Superantigens induced Tcell activation	
3.1.4	T cell differentiation (Memory and Effector cells)	
3.2	Cell mediated effector response	
3.2.1	General properties of effector T cells	
3.2.2	Cytotoxic T cells and destruction of target cell by perforin /granzyme pathway and Fas pathway	03
3.2.3	Killing mechanism of NK cells	
3.2.4	Antibody mediated cell cytotoxicity (ADCC)	
3.3	B cell	
3.3.1	B cell receptor and co-receptor-structure and function	
3.3.2	Development and maturation of B cells	
3.3.3	B cell activation and Differentiation	
3.3.3.1	Thymus dependant and independent antigens	04
3.3.3.2	Signal transduction pathway activated by BCR-overview	
3.3.3.3	Role of T _H cell in B cell response-Formation of T-B conjugates,CD40/CD40L interaction ,T _H cells cytokine signals	
3.4	Humoral Response	
3.4.1	Primary and secondary responses	
3.4.2	In vivo sites for induction of Humoral response	03
3.4.3	Germinal centres and antigen induced B cell Differentiation	
3.4.3.1	Cellular events within germinal centres-Overview	
3.4.3.2	Affinity maturation, somatic hyper-mutation and class switching	
3.4.3.3	Generation of plasma cells and memory cells	
Sub Unit	Unit IV: Immunology – II	15 Lectures
4.1	Vaccines	
4.1.1	Active and passive immunization	

4.1.2	Types of vaccines–Inactivated and attenuated vaccines, Subunit vaccines(Toxoid vaccines, Polysaccharide vaccines, Recombinant antigen vaccines) , recombinant vector vaccines, DNA vaccines Use of adjuvants in vaccine	06
4.1.3	New vaccine strategies	
4.1.4	Ideal vaccine	
4.1.5	Route of vaccine administration, Vaccination schedule, failures in	
4.1.6	vaccination	
4.2	Immunohaematology	
4.2.1	Human blood group systems, ABO, secretors and non secretors, Bombay Blood Group, Rhesus system and list of other blood group systems	
4.2.2	Haemolytic disease of new born, Coombs test.	
4.2.3	Blood Transfusion, Cross matching, Transfusion reactions	
4.3	Hypersensitivity	04
4.3.1	Coombs and Gells classification	
4.3.2	Type 1 to Type 4 Hypersensitivity : Mechanism and manifestation	
4.4	Autoimmunity	02
4.4.1	Definitions of autoimmunity, Immune tolerance and Immune suppression	
4.4.2	Types of autoimmune diseases	
4.4.3	Proposed Mechanisms	
4.4.4	Treatment	
	Text books: 1. Jawetz, Melnick and Adelberg’s Medical Microbiology, 26 th edition, Lange publication 2. Ananthanarayan and Panicker’s, (2017) Textbook of Microbiology, 10 th edition , Universities Press 3. Prescott, Harley and Klein,(2011)“Microbiology”, 8 th edition , McGraw Hill International edition. 4. Kindt, Goldsby, Osborne (2007)Kuby Immunology,	

6th edition, W H Freeman and Company

5. Pathak & Palan, (2011) Immunology: Essential & Fundamental, 3rd edition, Capital Publishing Company

6. Fahim Khan, (2009) Elements of Immunology, Pearson Education

7. Robert Bauman, (2015), Microbiology with diseases by body system., 4th Edition, Pearson Education Limited

8. Patrick R. Murray, Ken S. Rosenthal, (2005), Medical Microbiology, Elsevier Mosby

Reference books / Internet references:

1. Baron Samuel, Medical Microbiology, 4th edition
<http://www.ncbi.nlm.nih.gov/books/NBK7627/>

2. Kuby Immunology, 7th edition, WH Freeman and Company
<http://www.macmillanlearning.com/catalog/static/whf/kuby/>

Course Code SMIC603	Course Title: MICROBIAL BIOCHEMISTRY PART II	2.5 Credits Lectures/Week 4
	Learning Objectives : <ul style="list-style-type: none"> ➤ To study metabolism of lipids, fatty acids, nucleotides and amino acids ➤ To learn catabolism of proteins and aliphatic hydrocarbons ➤ To understand metabolic regulation and photosynthesis To learn metabolism of nitrate, sulphate and lithotrophy	
	Learning Outcomes: On completion of this course the student will <ul style="list-style-type: none"> ➤ Have learnt about the degradation and biosynthesis of lipids, hydrocarbons, proteins and nucleic acids ➤ Be knowledgeable about inorganic metabolism, prokaryotic photosynthesis and metabolic regulation 	
	THEORY	
Sub Unit	Unit – I: Lipid Metabolism & Catabolism of Hydrocarbons	15 Lectures
1.1	Introduction to Lipids	02
1.1.1	Lipids –Definition, classification & functions	
1.1.2	Types and role of fatty acids found in bacteria	
1.1.3	Common phosphoglycerides in bacteria	
1.1.4	Action of lipases on triglycerides /tripalmitate	
1.2	Catabolism of Fatty Acids and PHB	05
1.2.1	Oxidation of saturated fatty acid by oxidation pathway	
1.2.2	Energetics of oxidation of Palmitic acid	
1.2.3	Oxidation of propionyl CoA by acrylyl-CoA pathway and methyl citrate pathway	
1.2.4	PHB as a food reserve and its degradation	
1.3	Anabolism of Fatty Acids & Lipids	04
1.3.1	Biosynthesis of straight chain even carbon saturated fatty	
1.3.2	acid (palmitic acid)	
1.3.3	Biosynthesis of phosphoglycerides in bacteria	
	Biosynthesis of PHB	

1.4	Catabolism of aliphatic and aromatic hydrocarbons	04
1.4.1	Organisms degrading aliphatic hydrocarbons	
1.4.2	Hydrocarbon uptake mechanisms	
1.4.3	Omega oxidation pathway-	
1.4.3.1	Pathway in <i>Corynebacterium</i> and yeast	
1.4.3.2	Pathway in <i>Pseudomonas</i> Composition and architecture of membrane	
1.4.4	Growth with aromatic compounds --- ortho and meta cleavage	
Sub Unit	Unit II: Metabolism of Proteins and Nucleic Acids	15 Lectures
2.1	Protein/ amino acid catabolism	06
2.1.1	Enzymatic degradation of proteins	
2.1.2	General reactions of amino acids catalyzed by	
2.1.2.1	Aminoacid decarboxylases	
2.1.2.2	Amino acid deaminases	
2.1.2.3	Amino acid transaminases	
2.1.2.4	Amino acid racemases	
2.1.3	Metabolic fate of amino acids – Glucogenic and ketogenic amino acids	
2.1.4	Fermentation of single amino acid - Glutamic acid by <i>Clostridium tetanomorphum</i>	
2.1.5	Fermentation of pair of amino acids –Stickland reaction	
2.2	Anabolism of amino acids	02
2.2.1	Schematic representation of amino acid families	
2.2.2.	Biosynthesis of aminoacids of Serine family (Serine,Glycine and Cysteine)	
2.3	Catabolism of Nucleotides	03
2.3.1	Degradation of purine nucleotides up to uric acid formation	
2.3.2	Salvage pathway for purine and pyrimidine nucleotides	
2.4	Biosynthesis of nucleotides	04

2.4.1	Nomenclature and structure of nucleotides	
2.4.2	Role of nucleotides (high energy triphosphates)	
2.4.3	Biosynthesis of pyrimidine nucleotides	
2.4.4	Biosynthesis of purine nucleotides	
2.4.5	Biosynthesis of deoxyribonucleotides	
Sub Unit	Unit III: Metabolic Regulation	15 Lectures
3.1	Definition of terms and major modes of regulation	02
3.2	Regulation of enzyme activity	05
3.2.1	Non covalent enzyme inhibition	
3.2.1.1	Allosteric enzymes and feedback inhibition	
3.2.1.2	Patterns of FBI, combined activation and inhibition	
3.2.2	Covalent modification of enzymes	
3.2.2.1	Monocyclic cascades	
3.2.2.2	Examples of covalent modification (without structures)	
3.2.2.3	Regulation of Glutamine synthetase	
3.3	DNA binding proteins and regulation of transcription by positive & negative control	04
3.3.1	DNA binding proteins	
3.3.2	Negative control of transcription: Repression and Induction	
3.3.3	Positive control of transcription: Maltose catabolism in <i>E. coli</i>	
3.4	Global regulatory mechanisms	02
3.4.1	Global control & catabolite repression	
3.4.2	Stringent response	
3.5	Regulation of EMP and TCA cycle- (Schematic and Regulation of Pyruvate dehydrogenase Complex)	02
Sub Unit	Unit IV: Prokaryotic Photosynthesis & Inorganic Metabolism	15 Lectures

<p>4.1</p> <p>4.1.1</p> <p>4.1.2</p> <p>4.1.3</p> <p>4.1.4</p>	<p>Photosynthesis</p> <p>Definition of terms in photosynthesis (light and dark reactions, Hill reaction and reagents, Photophosphorylation)</p> <p>Photosynthetic pigments</p> <p>Location of photochemical apparatus</p> <p>Photochemical generation of reductant</p>	<p>04</p>
<p>4.2</p> <p>4.2.1</p> <p>4.2.2</p> <p>4.2.3</p>	<p>Light reactions in:</p> <p>Purple photosynthetic bacteria</p> <p>Green Sulphur bacteria</p> <p>Cyanobacteria (with details)</p>	<p>03</p>
<p>4.3</p> <p>4.3.1</p> <p>4.3.2</p>	<p>Dark reaction</p> <p>Calvin Benson cycle</p> <p>Reductive TCA cycle</p>	<p>02</p>
<p>4.4</p> <p>4.4.1</p> <p>4.4.1.1</p> <p>4.4.1.2</p> <p>4.4.1.3</p> <p>4.4.1.4</p> <p>4.4.1.5</p> <p>4.4.2</p> <p>4.4.2.1</p> <p>4.4.2.2</p> <p>4.4.2.2</p> <p>4.4.2.3</p> <p>Textbooks:</p> <p>1.Stanier,R.Y.,M. Doudoroff and E.A.Adelberg (1988) General</p>	<p>Inorganic Metabolism</p> <p>Assimilatory pathways:</p> <p>Assimilation of nitrate</p> <p>Ammonia fixation –Glutamate dehydrogenase, Glutamine synthetase, GS-GOGAT, Carbamoyl phosphate synthetase</p> <p>Biological nitrogen fixation (Mechanism for N₂fixation and protection of nitrogenase)</p> <p>Assimilation of sulphate</p> <p>Dissimilatory pathways:</p> <p>Nitrate as an electron acceptor (Denitrification in <i>Paracoccus denitrificans</i>)</p> <p>Sulphate as an electron acceptor</p> <p>Lithotrophy – Enlist organisms and products formed during oxidation of Hydrogen, carbon monoxide , ammonia , nitrite, sulphur , Iron</p>	<p>03</p> <p>02</p> <p>01</p>

Microbiology, 5th edition, The Macmillan press Ltd.

2. Conn, E. E., P. K. Stumpf, G. Bruening and R. Y. Doi. (1987). Outlines of Biochemistry, 5th edition. John Wiley & Sons. New York.

3. Gottschalk, G., (1985), Bacterial Metabolism, 2nd edition, Springer Verlag

4. White, D., (1995), The Physiology and Biochemistry of Prokaryotes, 3rd edition, Oxford University Press

5. Nelson, D. L. and M. M. Cox (2005), Lehninger, Principles of biochemistry, 4th edition, W. H. Freeman and Company.

6. G. Moat, J. W. Foster, M. P. Spector. (2002), Microbial Physiology, 4th edition, WILEY-LISS

7. Madigan, M. T. and J. M. Martinko (2006). Brock Biology of Microorganisms. 11th edition, Pearson Prentice Hall.

Reference books:

1. Zubay, G. L. (1996), Biochemistry, 4th edition, Wm. C. Brown publishers

2. Zubay, G. L. (1996), Principles of Biochemistry, Wm. C. Brown publishers

3. D. Nelson and M. Cox (2008) Lehninger, Principles of Biochemistry, 5th edition, W. H. Freeman and Company

Course Code SMIC604	Course Title: BIOPROCESS TECHNOLOGY: PART-II	2.5Credits Lectures/Week: 4
	Learning Objectives <ul style="list-style-type: none"> ➤ To understand processes involved in fermentation of important products ➤ To gain knowledge of plant and animal tissue culture techniques ➤ To understand the salient features of quality management and regulatory procedures ➤ To understand working of instruments used in biochemical analysis 	
	Learning Outcomes: On completion of this course the student will <ul style="list-style-type: none"> ➤ Have understood the techniques used in animal and plant tissue , stem cells and their application and enzyme immobilization ➤ Get an insight into the basics of Pharmaceutical microbiology ➤ Know the principles and applications of Spectrophotometry, Flame photometry, Spectrofluorimetry and radioisotopes ➤ Have learnt the fermentation processes of some important fermentation products 	
	THEORY	
Sub Unit	Unit – I: Advances in Bioprocess Technology	15 Lectures
1.1	Animal Tissue Culture	05
1.1.1	Types of tissue culture and cell lines	
1.1.2	Applications of tissue culture	
1.1.3	Advantages and Limitations	
1.1.4	Equipments used	
1.1.5	Tissue culture media	
1.1.6	Protocols for routine characterization of cell lines (Viable cell count using haemocytometer)	
1.2	Stem cells	05
1.2.1	Stem Cell Biology	
1.2.2	Culturing Stem Cells	

1.2.2.1	Human ES , Human EG and Human EC	
1.2.3	Applications	
1.2.3.1	Therapeutic Cloning	
1.3	Plant tissue culture	
1.3.1	Introduction	
1.3.2	Requirements for in vitro culture, Methods of plant cells and tissue culture	05
1.3.3	Types of cultures of plant materials: explants, callus, organogenesis, root culture, shoot culture, micro propagation, suspension culture, protoplast culture, protoplast fusion and somatic hybridization.	
1.3.4	Application : production of disease resistant plants, production of virus free plant, In vitro selection of cell lines for disease resistance, micropropagation, secondary metabolites from cell culture,transgenic plants for crop improvement	
Sub Unit	Unit II: Pharmaceutical Microbiology	15 Lectures
2.1	Vaccine Preparation	03
2.2	Quality assurance and quality control	
2.2.1	Definitions, Chemical and pharmaceutical products	
2.2.2	Variables of batch process	07
2.2.3	Q.A and Q.C wrt.- Raw materials ,method of manufacturing, in process items, finished products, label and labeling, packaging materials	
2.2.4	Control of microbial contamination during manufacturing	
2.3	Sterilization control and assurance	02
2.4	Bioassay	
2.4.1	Introduction	03
2.4.2	Types: Diffusion, End Point, Turbidometric, Metabolic Response, Enzymatic	
Sub Unit	Unit III: Instrumentation and IPR	15 Lectures
3.3	Instrumentation: Principles, working and application of	
3.3.1	Spectrophotometry: UV ,Visible &IR	
3.3.2	AAS &AES (Flame photometry)	06

3.3.3	Spectrofluorimetry	
3.3.4	Radioisotopic Methods	
3.5	Intellectual property rights	
3.5.1	Genesis, Role of WTO and TRIPS	05
3.5.2	Overview of patent system	
3.5.3	Requirements for patentability	
3.5.4	Patent Categories	
3.5.5	Preliminary steps for patent applications	
3.5.6	Patent Procedures	
3.5.7	For biotech and microbiological products	
2.3	Immobilized enzyme and cells	
2.3.1	Introduction and Definitions	04
2.3.2	Methods	
2.3.3	Immobilized Enzyme Reactors	
2.3.4	Applications	
Sub Unit	Unit IV:Industrial Fermentations	15 Lectures
4.1	Penicillin and semisynthetic penicillins: Introduction, biosynthesis and regulation, strain development, production methods. Semisynthetic penicillins: Examples, production, advantages	03
4.2	Aminoglycoside: Streptomycin: Aminoglycoside antibiotics, biosynthesis, regulation of biosynthesis, strain development, production method, recovery.	03
4.3	Vitamin B₁₂: Occurrence and economic significance, structure, biosynthesis, production based on media containing carbohydrates by- <i>Propionibacteria</i> and <i>Pseudomonas</i> Recovery.	02

<p>4.4</p>	<p>Citric acid: Introduction, strains used for production, biosynthesis, nutrient media, production processes- surface and submerged, product recovery.</p>	<p>03</p>
<p>4.5</p>	<p>Glutamic acid: Production strains, biosynthesis, effect of permeability on production, conditions of manufacturing, production process and recovery.</p>	<p>02</p>
<p>4.6</p>	<p>Steroid Transformation</p>	<p>02</p>
<p>Textbooks</p> <ol style="list-style-type: none"> 1. Casida L.E., (2009) "Industrial Microbiology" Reprint, New Age International (P) Ltd, Publishers, New Delhi. 2. Stanbury P. F., Whitaker A. & Hall S. J., (1997), "Principles of Fermentation Technology", 2nd Edition, Aditya Books Pvt. Ltd, New Delhi. 3. Stanbury P.F., Whitaker A. & Hall S.J (2017) "Principles of Fermentation Technology" 3rd edition 4. H. K. Das., "Text book of Biotechnology", 2nd and 3rd edition. 5. R.C. Dubey S. Chand. (2010) A textbook of biotechnology 4th edition. 6. H.A. Modi, (2009). "Fermentation Technology" Vol. 1 & 2, Pointer Publications, India 7. Okafor Nduka (2007) "Modern Industrial Microbiology and Biotechnology", Science Publications Enfield, NH, USA. 8. Crueger W. and Crueger A. (2000) "Biotechnology - A Textbook of Industrial Microbiology", 2nd Edition, Panima Publishing Corporation, New Delhi. 10. Prescott and Dunn's (1982) "Industrial Microbiology" 4th edition, McMillan Publishers. 11. Veera kumara L. "Bioinstrumentation", MJP Publisher 12. Hugo and Russell Pharmaceutical Microbiology, 7th edition, Blackwell Science. <p>Reference books</p> <ol style="list-style-type: none"> 1. Peppler, H.J. and Perlman, D. (1979), 		

	<p>"Microbial Technology". Vol 1 & 2, Academic Press.</p> <p>2. Williams, Bryan L; Wilson, 2nd edition." A Biologist's guide to principles and techniques of practical biochemistry" Baltimore: University Park Press, 1981.</p> <p>3. Wilson, Keith, 1936-; Goulding, Kenneth H, A Biologist's guide to principles and techniques of practical biochemistry 3rd edition., "London; Baltimore: E. Arnold, 1986.</p> <p>4. Wilson and Walker, (2008) "Principles and techniques of practical biochemistry" 5th edition. New Delhi, Cambridge University Press.</p>	
--	--	--



Semester VI – Practical

Course Code SMIC6PR1	PRACTICAL – I	3 Credits
	<p>Learning Objectives:</p> <ul style="list-style-type: none"> ➤ Develop soft skills ➤ To learn the practical aspects of immunohaematology and antibiotic sensitivity testing 	
	<ol style="list-style-type: none"> 1. Enrichment of coliphages, phage assay (pilot & proper). 2. Restriction digestion of lambda phage /any plasmid DNA 3. Beta galactosidase assay 4. Bioinformatics practicals 5. PCR (Demo) <p style="text-align: center;">On Line Practical</p> <ol style="list-style-type: none"> i. Visiting NCBI and EMBL websites & list services available, software tools available and databases maintained ii. Visiting & exploring various databases mentioned in syllabus and <ol style="list-style-type: none"> a. Using BLAST and FASTA for sequence analysis b. Fish out homologs for given specific sequences (by teacher – decide sequence of some relevance to their syllabus and related to some biological problem e.g evolution of some specific protein in bacteria, predicting function of an unknown protein from a new organism based on its homology) c. Six frame translation of given nucleotide sequence d. Restriction analysis of given nucleotide sequence e. Pair-wise alignment and multiple alignment of a given protein sequences f. Formation of phylogenetic tree <ol style="list-style-type: none"> 6. Animal cell culture (Demo) 7. Demonstration of malarial parasite in blood films 	

	<p>(Demo)</p> <ol style="list-style-type: none"> 8. Selection and testing of antibiotics using the Kirby-Bauer method 9. Susceptibility testing for antifungal agents 10. Determination of MBC of an antibiotic 11. E test 12. Blood grouping – Direct & Reverse typing 13. Determination of Isoagglutinin titer 14. Coomb's Direct test 15. Blood Transfusion : Compatibility Test 16. Demonstration experiments - VDRL, Rheumatoid Arthritis test 	
<p>Course Code SMIC6PR2</p>	<p>PRACTICAL – II</p>	<p>3 Credits</p>
	<p>Learning Objectives:</p> <ul style="list-style-type: none"> ➤ To learn estimations of biologically active compound ➤ Learning the principles and estimations of biocompounds 	
	<ol style="list-style-type: none"> 1. Detection of PHB producing bacteria 2. To study catabolite repression by diauxic growth curve. 3. Protein estimation by Lowry's method 4. Estimation of uric acid 5. Enrichment and isolation of Phenol degraders 6. Estimation of Phenol 7. Bioassay of an antibiotic (Ampicillin / Penicillin) 8. Bioassay of Cyanocobalamin. 9. Perform immobilization of yeast cells for invertase activity - making of beads, Determination of activity and count by haemocytometer and viable count. 10. Plant tissue culture – Callus culture (Demo). 11. Sterility testing of Pharmaceuticals : injectables and vaccines 12. Chemical estimation of Penicillin 13. Industrial Visit 	

Evaluation Scheme

[A] Evaluation scheme for Theory courses

I. Semester End Examination (SEE)- 100 Marks

[B] Evaluation scheme for Practical courses

1. Semester End Examination (SEE) – 100 Marks for Practical I and 100 Marks for Practical II.

